

TRANSFORMATION OF HEX-2-ENOPYRANOSYL TRICHLOROACETIMIDATES - A CORRECTION¹⁾

Ingolf Dyong[†], Hans Merten, and Joachim Thiem

Organisch-Chemisches Institut der Universität Münster,
Orléans-Ring 23, D-4400 Münster, Federal Republic of Germany

Summary: Contrary to previous results hex-2-enopyranosyl trichloroacetimidates do not rearrange to amino-branched glycals but give N-acylated hex-2-enopyranosyl amines.

Recently the [3.3]-sigmatropic rearrangement of allylic trichloroacetimidates²⁾ could be worked out into a general procedure for the synthesis of R-C-N-branched-chain amino sugars³⁾. This has been successfully confirmed by further studies with trichloroacetimidates of hex-2- and 3-enopyranosides as well as with sugar derivatives containing exocyclic allylic trichloroacetimidates^{3,4)}.

An extension of this approach to the 3-methyl-branched hex-2-enopyranosyl trichloroacetimidates should require particular interest. These could rearrange to yield amino-methyl-branched glycals which in turn comprise a reactive enol ether moiety and present attractive building units e.g. in N-iodosuccinimide glycosylation reactions⁵⁾.

By mild saponification of the corresponding methyl glycoside⁶⁾ 4-O-benzoyl-2,3,6-trideoxy-3-C-methyl- α/β -D-erythro-hex-2-enopyranose (3) was obtained. Treatment with sodium hydride and trichloroacetonitril at low temperature was anticipated to give the α - and β -trichloroacetimidates 4 which were reported to rearrange to the epimeric mixture of amino-branched chain glycals of ribo (5) and arabino configuration (6)³⁾. Further close inspection of this reaction revealed different results and demands a correction of the previous report³⁾.

[†] Deceased

In a low temperature n.m.r. experiment by treatment of 3 with the reagents the immediate formation of four trichloroacetimidates could be observed (four C=NH signals at δ 8.3 - 8.5 of different intensity). These are presumably both the α - and β -pyranose species 4 as well as their α - and β -furanose isomers 2. On heating to room temperature the intensity of these imino proton signals rapidly approached zero which points to a fast rearrangement reaction.

Thus the preparative reaction at -40°C and further heating to room temperature resulted in a mixture which after chromatography gave two crystalline isomers 8 α (57%) and 8 β (23%). They do not show significant differences in the optical rotation which renders the application of Hudson's isorotation rule⁷⁾ for the assignment of anomers impossible, as previously demonstrated in other hex-2-enopyranosyl derivatives⁸⁾. In ^1H n.m.r. the anomeric protons appear at similar shifts and each shows several small and one additional large $J(1,\text{NH})$ coupling⁹⁾.

Table

	Schmp. ($^{\circ}\text{C}$)	$[\alpha]_{\text{D}}^{20}$ (CHCl_3)	$\delta^{13}\text{C}(\text{C}-1)$	$J(\text{C}-1,\text{H})$	$\delta^{13}\text{C}(\text{C}-4)$	$\delta^1\text{H}(1-\text{H})$
<u>7α</u>	97	-18.0	86.94	167.4	89.15	6.39 dddq ^{a)}
<u>7β</u>	amorph	-29.2	88.24	167.1	88.82	6.47 dddq ^{b)}
<u>8α</u>	149	+86.6	74.27	163.3	72.19	5.84 ddddq ^{c)}
<u>8β</u>	144	+97.2	76.74	155.9	71.99	5.92 ddd ^{d)}

a) $J(1,\text{NH}) = 8.6$, $J(1,2) = 1.5$, $J(1,3-\text{CH}_3) = 1.5$, $J(1,4) = 1.5$;

b) $J(1,\text{NH}) = 8.7$, $J(1,2) = 1.3$, $J(1,3-\text{CH}_3) = 1.3$, $J(1,4) = 4.8$;

c) $J(1,\text{NH}) = 8.2$, $J(1,2) = 3.2$, $J(1,3-\text{CH}_3) = 1.5$, $J(1,4) = 1.6$, $J(1,5) = 0.3$;

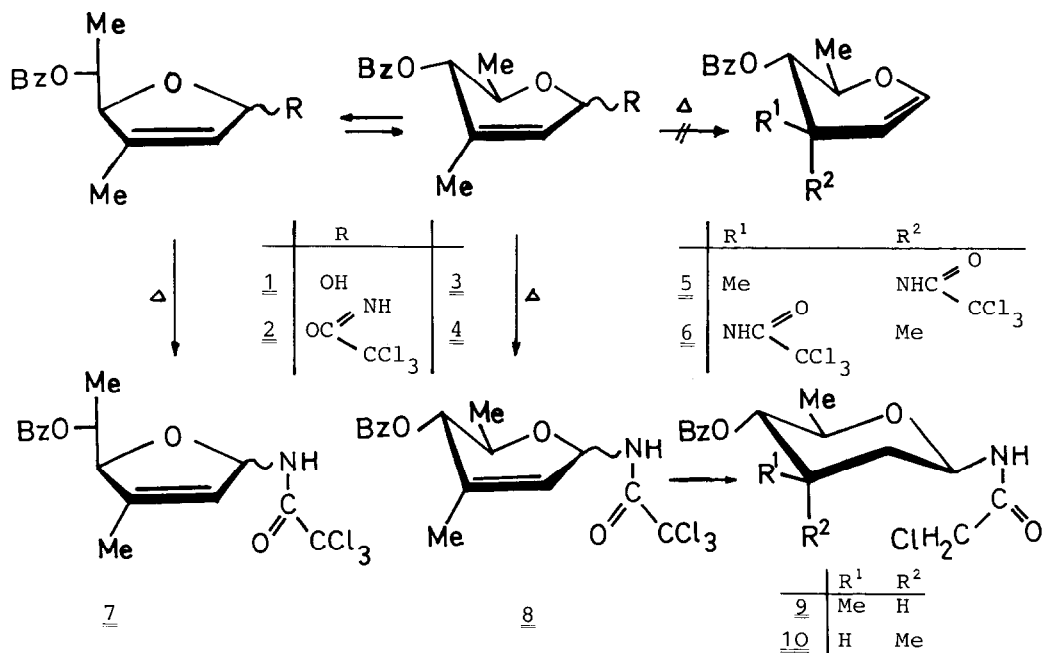
d) $J(1,\text{NH}) = 9.2$, $J(1,2) = 1.9$, $J(1,4) = 3.0$ Hz.

On account of these findings both the anomeric N-trichloroacetyl-4-O-benzoyl-2,3,6-trideoxy-3-C-methyl- α - (8 α) and β -D-erythro-hex-2-enopyranosyl amines (8 β) were obtained. The assignment of anomers is substantiated by carbon n.m.r. where 8 α shows an almost 8 Hz larger $J(\text{C}-1,\text{H})$ coupling than 8 β ¹⁰⁾. Further proof is provided by hydrogenolysis of the β anomer 8 β which results

in a mixture of (3R)- and (3S)-N-monochloroacetyl-4-O-benzoyl-2,3,6-trideoxy-3-C-methyl- β -D-erythro-hexopyranosyl amine [9 and 10, 72%, 9 : 10 = 5 : 2; 9 : 1-H δ = 5.32 ddd, $J(1, \text{NH}) = 8.8$, $J(1, 2a) = 10.9$, $J(1, 2e) = 2.2$ Hz].

When in contrast the same reaction is performed at -15°C with subsequent warming to room temperature a mixture of four isomers is obtained which were chromatographically separated (silica gel, CH_2Cl_2) and eluted as follows: 8 β (16%), 7 β (17%), 8 α (25%), and 7 α (30%). Again the assignment of the crystalline N-trichloroacetyl-5-O-benzoyl-2,3,6-trideoxy-3-C-methyl- α - (7 α) and the amorphous β -D-erythro-hex-2-enofuranosyl amines (7 β) cannot be based on the optical rotation or ^1H n.m.r. data but is supported by the considerable downfield shift of the C-4 carbons (Tab.). The carbon shifts of C-1 are consistent with the furanose structure, and because the $J(\text{C}-1, \text{H})$ coupling is virtually identical as could be expected from the almost planar system the anomeric assignment is tentative. Unfortunately both hex-2-enofuranosyl amines are labile and degraded in hydrogenation experiments.

These results lead to a tentative interpretation of the reaction course. Obviously depending on the exact conditions there can occur a preceding isomerisation of the hex-2-enopyranose 3 into the furanose isomer 1 under benzoate migration, probable mediated by sodium hydride.



Thus subsequent formation of all anomeric trichloroacetimidates 2 and 4 could be demonstrated, whereas a careful search including trapping conditions for an anticipated intermediate occurrence of glycols 5 and 6 did not meet with success. Under mild thermal conditions (below or maximal room temperature) cleavage of the imidates gives ion pairs of a glycosyl cation and an ambident trichloroacetimidate/trichloroacetamide anion. Their preferred recombination may be understood following Pearson's concept of a favourable interaction between the allylic oxocarbenium cation (hard acid) and the amide (hard base) which leads exclusively to the observed glycosyl amines 7 and 8 and no [3.3]-sigmatropic rearrangement products.

Acknowledgement: Support of this research by the Fonds der Chemischen Industrie and the Landesamt für Forschung des Landes Nordrhein-Westfalen is gratefully acknowledged.

References

- 1) 31. Communication on Biologically important Carbohydrates;
30. Communication: I. Dyong, H. Friege, and Th. zu Höne, Chem.Ber. 115, 256 (1982).
- 2) L.E. Overman, J.Am.Chem.Soc. 98, 2901 (1976).
- 3) I. Dyong, J. Weigand, and H. Merten, Tetrahedron Lett. 22, 2965 (1981).
- 4) J. Weigand, Diss. Univ. Münster 1982;
I. Dyong, J. Weigand, and J. Thiem, in preparation.
- 5) J. Thiem, H. Karl, and J. Schwentner, Synthesis 1978, 696;
J. Thiem and J. Elvers, Chem.Ber. 114, 1442 (1981).
- 6) I. Dyong and G. Schulte, Chem.Ber. 114, 1484 (1981).
- 7) C.S. Hudson, J.Am.Chem.Soc. 31, 66 (1909).
- 8) B. Coxon, H.J. Jennings, and K.A. McLauchlan, Tetrahedron 23, 2395 (1967).
- 9) R.H. Hall, A. Jordaan, and O.G. de Villiers, J.Chem.Soc., Perkin Trans. 1 1975, 626.
- 10) K. Bock and C. Pedersen, J.Chem.Soc., Perkin Trans. 2 1974, 293.

(Received in Germany 29 September 1983)